

TECHNICAL PROTOCOL
FOR
FRT-cm-FRT

FRT flanked,
Chloramphenicol Selection Cassette

(A006)

CONTENTS

1 Eppendorf tubes + manual

1. FRT-cm-FRT: PCR template (50 ng/ μ l, 20 μ l)
2. This manual

Store tube at -20°C

Please read

The products listed in this manual are for research purposes only. They are not designed for diagnostic or therapeutic use in humans, animals or plants. The Red[®]/ET[®] recombination technology is the intellectual property of Gene Bridges GmbH.

Conditions of use

3.1 Purchaser will not manufacture, copy, reproduce, transmit, distribute, sell, lease, transfer, or improve upon the MATERIALS without prior written consent from GENE BRIDGES.

3.2 All MATERIALS related to the technology must be purchased from GENE BRIDGES or its authorized distributors. Use of any of the stated products from a source other than GENE BRIDGES will exempt GENE BRIDGES from any and all liabilities and warranties.

3.3 All MATERIALS purchased by research organizations, universities and other non-profit organizations may not be used for any commercial purpose. These MATERIALS are to be used for research purposes only. The MATERIALS may not be used to provide a commercial or non-commercial service, of any kind.

3.4 The purchase of MATERIALS by a private consumer is neither intended nor permitted.

Short Description:

“FRT-cm-FRT” cassette is designed to allow chloramphenicol selection in prokaryotic cells.

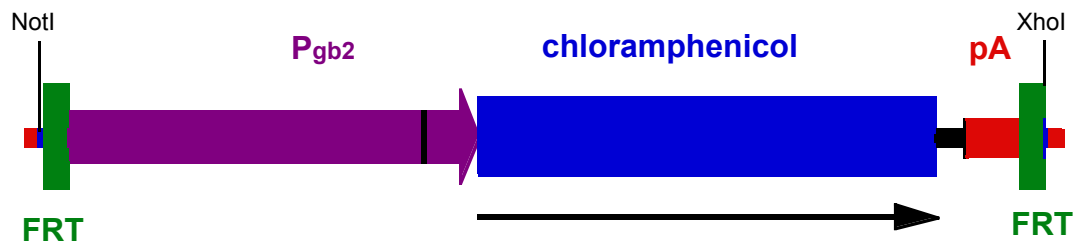
The prokaryotic promoter gb2 driving the gene for chloramphenicol resistance is a slightly modified version of the Em7 promoter; it mediates higher transcription efficiency than the generally used Tn5 promoter. A synthetic polyadenylation signal terminates the chloramphenicol expression. The cassette is flanked by FRT sites for later excision by Flp-recombinase. Unique *NotI* and *XhoI* sites flank the cassette for convenient cloning with restriction sites.

Using the provided PCR template one can easily create a FRT-cm-FRT cassette flanked by any other restriction sites to clone the cassette into the vector of choice. The restriction sites can be introduced by adding the corresponding sequence in the PCR primer.

The template can easily be used to engineer the *E. coli* genome by Red[®]/ET[®] Recombination.

The “FRT-cm-FRT cassette” is not linear but plasmid based (3302 bp in size). Due to its R6K origin the plasmid cannot replicate in most *E. coli* strains. The PCR product can therefore be used directly for downstream applications without any further purification.

At least 20 PCR reactions can be performed using 1 µl per reaction as template.



NotI

```

1  AATTAACCTCACTAAAGG GCGGCCG GAAAGTTCCTATTCTCTAGAAAGTATAGGAACTTC ATTCTACCGG
72  GTAGGGGAGG CGCTTTTCCC AAGGCAGTCT GGAGCATGCG CTTTAGCAGC CCCGCTGGGC ACTTGGCGCT
142 ACACAAGTGG CCTCTGGCCT CGCACACATT CCACATCCAC CGGTAGGCGC CAACCGGCTC CGTTC'TTGG
212 TGGCCCTTC GCGCCACCTT CCACTCTCC CCTAGTCAGG AAGTTCCTCC CCGCCCGCA GCTCGCGTCG
282 TGCAGGACGT GACAAATGGA AGTAGCACGT CTCACTAGTC TCGTGCAGAT GGACAGCACC GCTGAGCAAT
352 GGAAGCGGT AGGCC'TTGG GGCAGCGGCC AATAGCAGCT TTGCTCCTTC GCTTCTGGG CTCAGAGCT
422 GGAAGCGGT GGTCCGGG GCGGGCTCAG GGGCGGGCTC AGGGCGGGG CGGGCGCCG AAGTCTCTCC
492 GGAGGCCCG CATTCTGCAC GCTTCAAAG CGCACGCTG CCGCGCTGTT CTCCTCTTC TCATCTCCGG
562 GC'TTTCGAC C TGCAGC AGCACGTGTT GACAATTAAT CATCGGCATA GTATATCGGC ATAGTATAAT
629 ACGACAAGGT GAGGAACTAA ACC ATG GAG AAA AAA ATC ACT GGA TAT ACC ACC GTT GAT ATA TCC
      1 Met Glu Lys Lys Ile Thr Gly Tyr Thr Thr Val Asp Ile Ser
694 CAA TGG CAT CGT AAA GAA CAT TTT GAG GCA TTT CAG TCA GTT GCT CAA TGT ACC TAT AAC CAG
15 Gln Trp His Arg Lys Glu His Phe Glu Ala Phe Gln Ser Val Ala Gln Cys Thr Tyr Asn Gln
757 ACC GTT CAG CTG GAT ATT ACG GCC TTT TTA AAG ACC GTA AAG AAA AAT AAG CAC AAG TTT TAT
36 Thr Val Gln Leu Asp Ile Thr Ala Phe Leu Lys Thr Val Lys Lys Asn Lys His Lys Phe Tyr
820 CCG GCC TTT ATT CAC ATT CTT GCC CGC CTG ATG AAT GCT CAT CCG GAA TTC CGT ATG GCA ATG
57 Pro Ala Phe Ile His Ile Leu Ala Arg Leu Met Asn Ala His Pro Glu Phe Arg Met Ala Met
883 AAA GAC GGT GAG CTG GTG ATA TGG GAT AGT GTT CAC CCT TGT TAC ACC GTT TTC CAT GAG CAA
78 Lys Asp Gly Glu Leu Val Ile Trp Asp Ser Val His Pro Cys Tyr Thr Val Phe His Glu Gln
946 ACT GAA ACG TTT TCA TCG CTC TGG AGT GAA TAC CAC GAC GAT TTC CGG CAG TTT CTA CAC ATA
99 Thr Glu Thr Phe Ser Ser Leu Trp Ser Glu Tyr His Asp Asp Phe Arg Gln Phe Leu His Ile
1009 TAT TCG CAA GAT GTG GCG TGT TAC GGT GAA AAC CTG GCC TAT TTC CCT AAA GGG TTT ATT GAG
120 Tyr Ser Gln Asp Val Ala Cys Tyr Gly Glu Asn Leu Ala Tyr Phe Pro Lys Gly Phe Ile Glu
1072 AAT ATG TTT TTC GTC TCA GCC AAT CCC TGG GTG AGT TTC ACC AGT TTT GAT TTA AAC GTG GCC
141 Asn Met Phe Phe Val Ser Ala Asn Pro Trp Val Ser Phe Thr Ser Phe Asp Leu Asn Val Ala
1135 AAT ATG GAC AAC TTC TTC GCC CCC GTT TTC ACC ATG GGC AAA TAT TAT ACG CAA GGC GAC AAG
162 Asn Met Asp Asn Phe Phe Ala Pro Val Phe Thr Met Gly Lys Tyr Tyr Thr Gln Gly Asp Lys
1198 GTG CTG ATG CCG CTG GCG ATT CAG GTT CAT CAT GCC GTT TGT GAT GGC TTC CAT GTC GGC AGA
183 Val Leu Met Pro Leu Ala Ile Gln Val His His Ala Val Cys Asp Gly Phe His Val Gly Arg
1261 ATG CTT AAT GAA TTA CAA CAG TAC TGC GAT GAG TGG CAG GGC GGG GCG TAAGCGGACTCTGGGGTT
204 Met Leu Asn Glu Leu Gln Gln Tyr Cys Asp Glu Trp Gln Gly Gly Ala ...
1328 CGAATAAAGACCACCAAGCGAC GTC TGA GAGTCCCTG CGGAATTCGG TACCAATAAA AGAGCTTTAT
      XhoI
1397 TTTCATGATC TGTGTGTGG TTTTGTGTG CGGCGC GAAAGTTCCTATTCTCTAGAAAGTATAGGAACTTC C TCGAG
1474 CCCTATAGTGAGTCGTATTA

```

Please take into consideration that the sequence given above does not reflect the complete plasmid but refers to the functional cassette.